

In Vitro Effects of Ethanolic Extract and Crude Alkaloids of *Prosopis farcta* Leaves on the Viability of *Echinococcus granulosus* Protoscolices in Comparison to Mebendazole

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ABSTRACT

The scolicidal effects of crude ethanolic extract and crude alkaloids of *Prosopis farcta* leaves on protoscolices of *Echinococcus granulosus* were appraised *in vitro*, in comparison with mebendazole (the drug of choice in the treatment of echinococcosis) at 37C⁰ and 4C⁰ and at different time intervals.

Both extracts exerted higher effect at 37C⁰, and the crude alkaloids were more effective than ethanolic extract as it gave a scolicidal effect especially at the lower concentrations used (62.5, 31.2 mg/ml) within shorter periods (3 days and 13 days at 37C⁰ and 4C⁰ respectively)

Prosopis farcta

Echinococcus granulosus

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INTRODUCTION

Hydatidosis is a parasitic infection caused by larval stage of the tapeworm *Echinococcus granulosus*. Adult worms inhabit the small intestine of carnivores, larval stage (hydatid cyst) inhabits tissues of herbivores and omnivores host including human.

Infective stage to man is the eggs (Zeibig, 1997). Although surgery is still the treatment of choice for operable cystic disease due to *Echinococcus granulosus*, chemotherapy with benzimidazole, such as mebendazole and albendazole, which may be of value prior to surgery and in inoperable cases (WHO, 1996).

Success of treatment with mebendazole depends highly on the localization of hydatid cyst. Akhan et al. (1994) showed that the cure percentage of pulmonary cysts using mebendazole was 2%, while it was 7% in case of hepatic hydatidosis. (Aidan and Yalin, 1996) reported that 35% of desiccated hydatid cysts from patients who were pretreated with mebendazole were non-fertile. Kuro et al. (1997) recorded those 30% cure percentage in patients with hepatic hydatidosis who were treated with albendazole and mebendazole. Hence, great efforts are being spent to find an effective medical cure for the disease.

Recently, many workers are focusing their researches toward the use of natural products in the treatment of disease including echinococcosis. Ilammo (2002) reported that the aqueous extracts of *Matricaria camomilla* and *Cyperus rotundus* possess an inhibitory effect on the viability of *Echinococcus granulosus* protoscolices. Mahmoud (2002) studied the effect of three plants on the viability of *Echinococcus granulosus* Protoscolices, which are *Thymus villosus*, *Cyperus rotundus* and *Peganum harmala*, respectively. She showed that the aqueous, ethanolic and alkaloid extracts of these plants have a good inhibitory effect on the viability of Protoscolices. Thus, the present work aims at studying the effects of ethanolic and alkaloidal extracts of *Prosopis farcta* leaves on the viability of *Echinococcus granulosus* Protoscolices.

Prosopis farcta is a member of Mimosaceae family. It is a struggling shrub with 30-100cm height, grows abundantly in Iraq (Rechinger, 1964). The genus *Prosopis* has been used in folk medicine as an astringent in rheumatic disorders and as a remedy against scorpion sting and snakebite. The leaves of *Prosopis spp.* were reported to relieve irritated conjunctiva as used by Indians in South Western U.S. and in Mexico (Usmanghani et al., 1982, and Aqeel et al., 1991) *Prosopis spp.* were also known to have medicinal properties. It was proved to have cytotoxic effects in its fruits, which exhibit significant activity against lung carcinoma (Ahmad and Sultana, 1989).

Juliforin, which is the main alkaloid of *Prosopis juliflora*, has been shown to possess antidermatophytic and antibacterial activities (Al-Shaikh Hamed and Al-Jammas, 1999). Usmanghani et al. (1982) isolated juliprosopin alkaloid from *Prosopis farcta* leaves, and then she studied its activity against some species of bacteria.

MATERIALS AND METHODS

Plant Materials:

Leaves of *Prosopis farcta* were collected from Mosul and its surrounding fields during June and August 2001. The plant was air-dried and the dried leaves were used. Biology department, College of Science, Mosul University, authenticated this plant.

Preparation of Ethanolic Extract:

The dried leaves were ground into coarse powder. Then extracted with 80% ethanol and filtered. The filtrate was evaporated using rotary vacuum evaporator. It was then lyophilized to get dry powder of the extract (Verport et al., 1988).

Preparation of Crude Alkaloids:

The powdered leaves were extracted with acid alcohol (1 N HCl in 80% ethanol) then pigments and unwanted materials were removed by shaking with petroleum ether. The water layer was alkalized with ammonium hydroxide. The precipitated alkaloids then were separated by filtration (Evans, 1997).

Protoscolices Collection and Suspension:

In this study, hepatic hydatid cysts of infected sheep were obtained from the municipal abattoir in Mosul City, Iraq, washed in several changes of sterile phosphate buffer saline (pH 7.2). Protoscolices were then suspended in sterile hydatid fluid ultrafiltered using 0.4 micropore filters, 2% of dimethyl sulphoxide was added to the fluid as organic solvent (Farjou and Al-Hussainawi, 1984). Two ml of the suspension containing 500 Protoscolices were transferred into each siliconized test tube.

The viability of the Protoscolices used was $\geq 90\%$. This was determined by peristaltic movement of protoscolices and negative staining with 0.1% aqueous eosin and flame cell movements (Smyth and Barrett, 1980).

The Effect of Ethanolic Extract on the Viability of the Protoscolices:

Two groups of Protoscolices suspensions were used. Each group consists of nine test tubes; the first was considered as control, the second was treated with 0.1 mg mebendazole/ml of the suspension. The remaining tubes (3-9) were treated with different concentrations of the ethanolic extract (500, 250, 125, 62.5, 31.2) mg of the extract/ml of the protoscolices suspension, respectively.

The first group was incubated at 37°C and the second group was incubated at 4°C. The viability of Protoscolices was examined for each group after each exposure period (each treatment was carried out in triplicates).

The effect of Crude Alkaloids on the Viability of the Protoscolices:

To prepare different concentrations of crude alkaloids extract, the same procedure in preparing ethanolic extract treatments was used as mentioned before.

Statistical Analysis used in this study includes F-test using analysis of variance (ANOVA table) and Dunnett's multiple range tests. The results were evaluated at $p < 0.05$ and $p < 0.01$ (Al-Rawi and Kalaf Allah, 1980).

RESULTS AND DISCUSSION

The antiparasitic activity of ethanolic extract and crude alkaloids of medicinal plant *Prosopis lareta* has been tested against *Echinococcus granulosus* Protoscolices *in vitro*, the study was carried out at 37°C and 4°C respectively (table I and table 2). ANOVA table (Table 3) showed that there are no significant differences among the inhibitory effect of all treatments (A) which are used in this study, but the interaction between treatments and exposure periods (A*P) showed high significant differences among viability percentage of the Protoscolices. This result means that the inhibitory effect of both extracts at both temperatures was weak (non-significant) and it got stronger (significant) with process of time. The delayed inhibitory effect of this plant may be due to the presence of small amounts of active constituents, i.e. scolicidal materials in the crude extracts which is used in the present study.

Table (3) also showed that there are high significant differences in the inhibitory effect of the extracts between the two temperatures used (B). Furthermore, interaction between the effect of temperatures and exposure periods (B*P) showed a high significant differences between viability percentage of Protoscolices. Comparing the mean viability percentages of Protoscolices at 37°C (table I) and at 4°C (table 2) it was evident that the inhibitory effect of both extracts used was higher at 37°C than at 4°C, and this result became more clear with the passage of time. This result is coincided with our former study in which the scolicidal effect of *Peganum harmala* seeds against *Echinococcus granulosus* protoscolices was stronger at 37°C than at 4°C (Hammoshi et al., 2002) This may be due to the fact that living cells are metabolically more active at 37°C (Al-Habib, 1991) and many of the inhibitory substances are more effective at this temperature.

Simultaneously, the survival period for Protoscolices incubated at 37°C is shorter than those incubated at 4°C, because of the faster autolytic activity of Protoscolices at 37°C (Andersen and Loveless, 1978; Mahmood and Al-Hannon, 1981). Hammoshi and Rahmo 2002 showed that incubation of *Echinococcus granulosus* protoscolices at 4°C *in vitro* keeps viability of protoscolices up to 18 days, while incubation of protoscolices at 37°C leads to destruction of the protoscolices within 3 days. They also demonstrated that vesiculation and evagination of protoscolices occurred within few hours after incubation at 37°C while they occurred at a much lower level after two days of incubation at 4°C. Thus, metabolic activities of protoscolices are increased at 37°C including the autolytic activity) leading to faster destruction of the protoscolices, but better reaction with metronidazole and other tested extracts and *vice-versa* when the protoscolices are incubated at 4°C (Table 1 and Table 2).

Table 1 : The effect of ethanolic extracts and crude alkaloids of *Prosopis farcta* leaves on the viability % of *Echinococcus granulosus* protoscolices *in vitro*, 37 C⁰ in comparison with mebendazole and control groups.

Treatments	Concentration	Mean viability% of Protoscolices after...day			
	Mg/ml	1st	2nd	3rd	5th
*Control	0	99	91	85	3
Mebendazole	0.1	9	0	0	0
Ethanolic extract	500	75	37	0	0
	250	88	67	0	0
	125	93	66	10	0
	62.5	95	68	9	0
	31.2	95	81	10	0
Crude alkaloids	500	43	26	0	0
	250	56	38	0	0
	125	81	33	0	0
	62.5	82	64	0	0
	31.2	92	58	0	0

N: 3 replicates

*Control group=500 potoscolices/ml of hydatid fluid.
Percentage at zero time considered as 100%

Table 2 : The effect of ethanolic extracts and crude alkaloids of *Prosopis farcta* leaves on The viability % of *Echinococcus granulosus* protoscolices *in vitro*, 37 C⁰ in comparison with mebendazole and control groups.

Treatments	Concentration	Mean viability% of Protoscolices after...day								
	mg/ml	1st	2nd	3rd	5th	7th	9th	11 th	13th	15th
*Control	0	98	90	81	73	62	44	30	7	2
Mebendazole	0.1	20	3	0	0	0	0	0	0	0
Ethanolic extract	500	70	53	21	0	0	0	0	0	0
	250	85	77	62	50	33	10	2	0	0
	125	92	72	58	56	52	35	26	7	0
	62.5	96	91	82	78	70	37	28	8	0
	31.2	95	88	88	80	67	40	33	12	0
Crude alkaloids	500	46	25	14	0	0	0	0	0	0
	250	82	67	45	24	10	3	0	0	0
	125	90	78	72	70	52	34	20	0	0
	62.5	97	91	90	75	50	32	25	2	0
	31.2	92	92	90	83	64	45	31	1	0

N: 3 replicates

*Control group=500 potoscolices/ml of hydatid fluid.
Percentage at zero time considered as 100%.

Table 3 : Combined ANOVA table for the effect of treatment (A), temperature (B) and exposure times (P) on the viability of *Echinococcus granulosus* protoscolices *in vitro*.

Source of variance	Degree of freedom	Sum of square	Mean square	Calculated F
A	11	63068.27	5733.48	0.78
B	1	61302.707	61302.707	**8.11
AxB	11	1939.583	1763.05	0.24
Error a	46	335317.17	7289.503	
P	8	270779.24	33847.405	**631.8
AP	88	384255.12	4366.54	**81.51
BP	8	8404.16	1051.02	**19.62
AbP	88	3685.767	41.88	0.78
Error b	384	20571.27	53.57	
Total	647	527044.61		

**P<0.01

In table 4, the results were arranged from highest to lowest inhibitory effect on the viability percentage of the *Protoscolices* at 37°C (using capital letters). Table 1 shows that the *Protoscolices* treated with both extracts giving a progressive decline in survival means and this is marked in case of ethanolic extract since the first two days of incubation, especially at the high concentrations 500 and 250mg/ml, while inhibitory effect of the crude alkaloids was evident from the highest to the lowest concentration used in this research. Hence all *Protoscolices* were killed before the third day of incubation.

Pursuing levels of effects for each treatment (Table 4) show that both ethanolic extract and crude alkaloids exhibit less inhibitory effect than mebendazole. But generally the scolicidal effect of crude alkaloids was stronger than that of ethanolic extract.

Observing mean viability percentage of *Protoscolices* at 4°C (Table 2) shows that both ethanolic and alkaloidal extract possessed low scolicidal effect in comparing with mebendazole. They also exhibit weak or no inhibitory effect in the low concentrations (G2.5,31.2mg/ml). But when you follow up levels of inhibitory effect of each treatment at this temperature (Table 5), it seems that the crude alkaloids have more potent scolicidal effect than ethanolic extract. It appears that the alkaloids used in the present study rather have a good scolicidal effect, even present at low concentrations in the crude extract of *prosopisfarcta* leaves.

The use of some drugs is probably limited due to the risk of their side effects and toxicity. Plants are relatively safe and rich source of therapeutic compound, and alkaloids in certain respect rank among the most significant naturally occurring substances (Aqeel et al. 1991) Typical alkaloids were derived from plant source sharing an important location in chemotherapy world; Alkaloids found in all parts of the plant or concentrated in certain parts (Kotb, 1981).

Table 4 : Leaves of the inhibitory effects of ethanolic and alkaloidal extracts on the viability of *Echinococcus granulosus* protoscolices at 37°C⁰ in comparing with mebendazole and control group (Duncan's test at P>0.05)

Treatments	Concentration Mg/ml	Viability mean of Protoscolices	Level of effect
Control	0	694.86	H
Mebendazole	0.1	58.38	A
Ethanolic extract	500	297.66	BC
	250	376.68	CDE
	125	448.62	EFG
	62.5	454.77	FG
	31.2	483.54	H
	Crude alkaloids	500	221.995
250		264.43	BC
125		302.91	CD
62.5		393.54	DEF
31.2		374.76	CDE

*Each value represents the mean of three replicates. *There are no significant differences between the values that subscribed by one litter or more.

Table 5 : Leaves of the inhibitory effects of ethanolic and alkaloidal extracts on the viability of *Echinococcus granulosus* protoscolices at 4C⁰ in comparing with mebendazole and control group (Duncan's test at P>0.05) at P>0.05).

Treatments	Concentration Mg/ml	Viability mean of Protoscolices	Level of effect
Control	0	1255	HI
Mebendazole	0.1	114.87	A
Ethanolic extract	500	396.87	C
	250	862.86	E
	125	1074.02	F
	62.5	1262.04	G
	31.2	1284	HI
Crude alkaloids	500	288.54	B
	250	660.74	D
	125	1071.88	F
	62.5	1205.81	GH
	31.2	1296.35	I

*Each value represents the mean of three replicates. *There are no significant differences between the values that subscripted by one letter or more.

Mahmoud, (2002) reported that the crude alkaloids of *Thymus vulgaris* leaves,

Cypenls rotundus tubercles and *Peganum hannala* seeds, exhibited the best scolicidal effect in comparison with aqueous and ethanolic extracts of the same plants. Hammoshi et al. (2002). also showed that the crude alkaloids of *Peganlln hannala* seeds were more effective than the ethellolic extract against *Echchyinococcus granulosus* protoscolices.

Several alkaloids have been derived from leaves of *Prosopis* spp like spcigerine julillorine, jlllfloricine, jlllloridine and juliprisopine that were reported to have medicined value (Ahmad et al., 1979 ; Ahmad and Qazi, 1985). Ahmad and Sultana (1989) showed that the aqueous and alco_1O1ic extracts of *Prosopisjuli./dra* have an antibacterial activity. Usmanghani et al., (1982).Isolated two alkaloids from shade dried leaves of *Prosopis glandulosa* and *Prosopis.!arcta*, and characterized them as juliforine and juliprosopine. They also found that the alcoholic extract and isolated juliprosopine showed antibacterial activity against *Bacillus subtilis*, *Bacillus magatherium* and *Sarcinnia lulea* .Juliflorine had been shown to possess antibacterial, antidermatophytic and non-mutagenic activity (Aqeel et al., 1991) .

However, further investigations are needed to isolate and purify the different types of alkaloids from the crude alkaloid of *Prosopis fareta* leaves and then identify the scolicidal effect of each.

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REFERENCES

- Ahmad, V.U. and Qazi, S. (1985). Studies on the structure of julilloricine. J. of Chem. Soc. Pale 1(4), pp.347-350.
- Ahmad, V. U .and Sultana, A. (1989). A Terpenoid diketone from the leaves of *Jatropha gossypifolia*. J. Phytochemistry. 28(1).289-278,
- Ahmad, V.U.; Usmanghani, K. and Saqib, Q.N. (1979). Occurrence of julilloridine in *Prosopis juliflora* Torr. SCI. Pharm. 47, 333-334.
- Aktan, O. and Yalin, R. (1996). Preoperative albendazole treatment for liver hydatid disease decreases the viability of the cyst. Eur.J. of Gastroenterology and Hepat. 8(9), 877-879.
- Aktan, O.; Ozmen, M. N.; Dincel, A.; Goemen, A. and Kalyoneu, E. (1994). Percutaneous treatment of pulmonary hydatid cysts. Cardio and Interventional Radiol 17,227-275.
- Al-Habib, O. A. (1991). Animal Physiology, Oar Al-Kotob for printing and Publication, Mosul, 112-140.
- Al-Hammo, R. N. (2002). The inhibitory effect of some plant extracts on the viability of *Protoscolices in vitro*. Accepted for publication in Raf. J. of Sci.2(14).
- Al-Rawi K. and Kalaf Allah, A.M. (1980). The Completely Randomized Design. in: Design and Analysis of Agricultural experiments. Dar Al-kotob for Printing and Publication, Mosul, 37-94.
- Al-Shaikh Hamed, W.M.A and Al-Jammas, M.A.A. (1999). The antimicrobial activity of alkaloidal fraction of *Prosopis Juliflora*. Iraqi J. Veterinarian Science. 2(12), 24-29
- Andersen, F.L. and Loveless, R.M.. (1978). Survival of *Protoscolices* of *Echinococcus granulosus* at constant temperatures. J. Parasitol.8278,(1)64 :
- Aqeel, A.; Khurshed, A.k and Viqaruddin, A. (1991). Toxicological studies of the antimicrobial alkaloid juliflorine. J. Arzneim. forsch/ Drug Research.41(I). Nr2., 151-154.
- Evans W.C. (1997). Treas and Evans Pharmacognosy. Fourth ed. W. L3. Saunders Company limited, V. P. India, 340-408.
- Faljou, I. B. and Al-Ilussainawi, S.S. (1984). Effect of mebendazole on the survival of hydatid *Protoscolices* of *Echinococcus granulosus* *in vitro* and *in vivo*. J. Fac. Med.Baghdad, 26. (4), 33-44.
- Hammoshi. M.II.; Sharee C. A.Y. and Younis, G.TH. (2002). Effect of ethanolic extract and crude alkaloides of *Peganum hanna* seeds on the viability of *Echinococcus granulosus* *Protoscolices in vitro*. Accepted for publication in Raf. J. of Sci.4(14).
- hammoshi, M.H. and Rahemo, L.F. (2002). The effect of temperature on viability of the *Protoscolices* in cysts of *Echinococcus granulosus* as tested *in vitro*. Raf.J. of Sci.13(13),49-57.
- Kotb, F. (1981). Medicinal Plants, their agriculture and content. Alkaloids. Al Marrek house, Egypt. 88-99 .
- Kuro, M. S.; Nazir, D. M.; Javid Jul Khan, B. A.; Chulam, N.; Yattoo, M. D.; Altaf, S.M.D. ,and Jellani, S. G. (1997). Percutaneous drainage compared with surgery for

- hepatic hydatid cysts. *The New Eng. J. Med.*, 337 (13), 881-887.
- Mahmoud. S.W. (2002). The effect of some plant extracts on the viability of *Echinococcus granulosus* protoscolices of human and sheep origin *in vitro* and their growth *in vitro*. M.Sc. Thesis, College of Education, University of Mosul.
- Mahmoud, S.S. and Al-Hanoon, Z.A., (1988). The effect of temperature on the survival of hydatid protoscolices/ *Journal of College of Veterinary Medicine, Mosul*. 2(1 and 2), 35-140.
- Rechinger, K.H. (1964). *Flora of Lowland Iraq*. Publisher in Weingheim, Germany, 205-230.
- Smyth, J. D. and Barrett, S. (1980). Procedures for testing the viability of human hydatid cysts following surgical removal especially after chemotherapy. *Trans R. Soc. Trop. Med. Hyg.* 74,649-652.
- Usmanghani, K.; Saqib, Q.N. and Ahmed, V.D. (1982). Occurrence of juliprosopine in *Prosopis glandulosa* Torr. And *Prosopis farcta* (Banks and Sol.) Macbride. *J.Chem.Sci.Pak.*, 4(4), 285-287.
- Verport, R.; Tginastoi, A.; Vandoorne, H. and Svendsen, A. B. (1988). Medicinal plant of Surinam. I-Antimicrobial activity of some medicinal plants. *J. Ethnopharmacol.* 5,221-226.
- WHO. (1996). Guidelines for treatment of cystic and alveolar echinococcosis in human. *Bulletin of the World Health Organization*. 74,p: 23]-242.
- Zeibig E.A. (1997). *Clinical Parasitology, The Cestodes*. W.B.Sauncier Company, U.S.A. 195-197